



Brazilian Journal of Physical Therapy

<https://www.journals.elsevier.com/brazilian-journal-of-physical-therapy>



ORIGINAL RESEARCH

Evaluation of the female pelvic floor with infrared thermography: a cross sectional study



Halana Karolina Vicente da Silva, Maria Clara Eugênia Oliveira, Edson Silva-Filho*,
Adriana Gomes Magalhães, Grasiela Nascimento Correia,
Maria Thereza Albuquerque Barbosa Cabral Micussi

Universidade Federal do Rio Grande do Norte, Natal, RN, Brazil

Received 12 February 2020; received in revised form 2 December 2021; accepted 5 January 2022
Available online 26 January 2022

KEYWORDS

Color;
Muscles;
Temperature;
Women;
Women's health

Abstract

Background: Infrared thermography (IRT) is an easy-to-use, noninvasive and pain-free tool that can be used to evaluate function of the pelvic floor (PF) muscles.

Objective: To analyze vaginal manometry, temperature, and percentage of colors achieved through IRT of the PF muscles at rest and during maximum voluntary contraction. The relationship between PF muscles strength and IRT temperature was also assessed.

Methods: Two-hundred and thirty-one women (mean \pm SD age: 58.4 ± 5.9 years) participated in this study. IRT recorded the minimum, average, and maximum temperatures, and the colors of the PF area at rest and during maximum voluntary contraction. The pressure applied during the three maximum voluntary PF contractions was evaluated through vaginal manometry.

Results: The women had a PF average temperature of $36.4 \pm 0.8^\circ\text{C}$. There were no differences in the IRT temperatures between rest and during PF muscles contraction. The percentages of white, red, orange, yellow, green, cyan, and blue colors were different at rest and during contraction. Warm colors became more visible in the center of the image during the PF muscles maximum voluntary contraction. There was a positive correlation between the PF average temperature and PF manometry ($r=0.7$; $p=0.001$).

Conclusion: The IRT was not able to detect differences in the temperature of the PF area between at rest and during contraction. However, a strong correlation between PF temperature and vaginal manometry was found.

© 2022 Associação Brasileira de Pesquisa e Pós-Graduação em Fisioterapia. Published by Elsevier España, S.L.U. All rights reserved.

Introduction

Pelvic floor (PF) dysfunction causes structural and functional changes and is associated with malfunctions and injuries of the pelvic floor muscles (PFM). The most frequent PF

* Corresponding author at: Rua Teodorico Bezerra, CEP 59200-000, Santa Cruz, Rio Grande do Norte, Brazil.

E-mail: meneses.edson@yahoo.com.br (E. Silva-Filho).

dysfunctions in women are urinary incontinence,¹ fecal incontinence,² and sexual problems.³ These dysfunctions have been associated with age, pregnancy, childbirth injuries, obesity, genetic factors, and lack of physical activity.^{1,2}

Evaluation of the PFM functions has an essential role in the diagnosis and treatment of PF dysfunctions.⁴ Digital palpation, manometry, electromyography, ultrasound, and magnetic resonance imaging are commonly used to assess PFM in clinical practice.⁵

Infrared thermography (IRT) is an imaging method used to assist in the diagnosis of selected neurological and musculoskeletal conditions, and in the early detection of conditions such as periprosthetic joint infections,⁶ breast cancer,⁷ musculoskeletal dysfunctions,⁸ and injuries.⁹ IRT measures the infrared radiation emitted by a surface and transforms the radiation intensity into temperature values. This technique has been demonstrated to have high sensitivity and specificity for clinical diagnoses of some conditions related to cancer and muscular impairments.^{10,11}

IRT has been used to explore temperature alterations in some systems of the human body.¹² Several PF problems have been associated with vascular alterations, such as those involved in sexual and some climacteric dysfunctions.¹³ Possibly, IRT may be able to detect malfunctioning of the PF in the very early stages, even before the onset of symptoms. One advantage over other techniques is that it can be used during periods that are not possible to perform intravaginal evaluations and is potentially more comfortable and better accepted by women. Similar to other studies that have associated tissue temperatures with muscular performance^{14,15} an evaluation of the PF by IRT may detect temperature patterns that might be associated with muscular dysfunction. In addition, IRT may be a relatively low-cost additional resource to assess PFM.

This study aimed to analyze vaginal manometry, temperature, and the percentages of various colors through IRT of the PF area in both the resting state and during maximum voluntary contraction. The relationship between PFM strength and the temperature of the PF area was also assessed.

Methods

Study design

This cross-sectional study followed the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) Statement recommendations.¹⁶ It was approved by the Research Ethics Committee from *Universidade Federal do Rio Grande do Norte*, Natal, RN, Brazil (number 2.328.269) and was conducted in accordance with the principles of the Helsinki Declaration. All participants signed the Informed Consent before participating in the study.

Participants and setting

Participants were recruited from February 2018 to February 2019 at the Gynecology and Climacteric outpatient clinics of the *Januário Cicco Maternity School* (Natal/RN/Brazil). The women were included if they met the following criteria: (1) age between 45 and 70 years; (2) nonpregnant; (3) no

urinary/vaginal infection; (4) without vasomotor disease or gynecological bleeding; (5) no gynecological surgery or childbirth in the last six months; and (6) no neurological disorders that could compromise the IRT assessment. Those who used vasomotor drugs, did not present any ability to contract the PFM (as detected by manual palpation), presented with pelvic organ prolapse (stage three or four according to the Baden-Walker scale),¹⁷ reported pain/discomfort during a vaginal probe introduction, or had a PFM contraction associated with activation of the abdominal, gluteal, and hip adductor muscles were excluded from the study.

To assess the PFM contraction, all women were in supine, with bent knees and flexed and abducted hips. They were asked to perform three voluntary PFM contraction attempts. An examiner with five years of experience in women's health physical therapy performed all the evaluations. The verbal command given by the examiner to the participants was to contract the PFM by squeezing it with maximum force as if they were holding their urine.

Variables and measurements

Initially, data related to age, marital status, parity, types of delivery, menstrual cycle, and body mass index were collected. Next, PF temperature and vaginal manometry were assessed in this order.

Thermography was measured using a Flir® thermographic camera, model one (FLIR Systems, Inc., Wilsonville, OR, USA) that was attached to an iPhone 6S (Apple, Inc., Cupertino, CA, USA). The device weighs 30 g and contains two cameras, a Lepton™ thermal sensor (160 × 120 pixels) and a visible VGA camera (640 × 480 pixels). The two images obtained from these cameras were merged using the Multi-Spectral Dynamic Imaging (MSX) technology, resulting in one thermal image with a resolution of 640 × 480 pixels. The FLIR One camera has a temperature capability ranging from -20°C to 120°C and can detect temperature differences as small as 0.1°C. The Flir® software was used to show the minimum, average, and maximum temperature.

Thermography was used to record the PFM at rest and during a single maximum voluntary contraction with a camera positioned perpendicular to the perineum region, which is the central point of a rectangular image 5 cm wide and 10 cm long. The images were analyzed using a rainbow chart.

Some factors were controlled to achieve surface thermography:¹⁸ room temperature between 18°C and 25°C, without lighting, with upward ventilation, 60% humidity as evaluated by a digital thermo-hygrometer. The distance between the camera and perineum region was 20 cm, and the patient acclimatization time to the setting before taking an image was 10 min.¹⁹ Those volunteers who had hair in the image collection region were shaved after agreeing to this procedure. The researcher used a razor blade and took care to not injure the skin. The evaluation always occurred between 8:00 AM and 11:00 AM, was conducted by the same trained evaluator (a women's health physical therapist with five years of experience in performing PFM assessments and IRT) and lasted for about 10 min.

The TermoCrivo® software package was used to illustrate the ratio of warm and cold colors based on the HSV model

(hue, saturation, and value). Recent studies showed good correlation between thermography and Doppler ultrasound in different diseases and body areas.²⁰⁻²² In this study, the colors were classified as warm if they displayed white, red, orange, or yellow and as cold if they showed green, cyan, blue, violet, or magenta.²³ For the color analysis using the software, the pixel count of each color was used, resulting in the acquisition of the percentage of white, red, orange, yellow, blue, cyan, green, violet, and magenta areas in the image. The total amount of pixels in each image was 307,200. The sum of the frequency of all colors reached 100%.

Lastly, the vaginal manometer Peritron® Model 9300V (Cardio Design, Australia) was used to measure the pressure produced by PFM contraction. All participants remained in the supine position and digital palpation was initially performed to confirm their ability to voluntarily contract the PFM. The vaginal probe was covered with a nonlubricated latex condom and then lubricated using a gel before introduction in the vagina. Next, the probe was introduced gently, without causing pain, approximately 3.5 cm into the vaginal cavity with the device turned off. A verbal command for a muscle contraction was given to hold the urine and squeeze the vagina with maximal force. The participants performed three maximum voluntary contractions, with a 30-s rest between the muscle contractions.²⁴ The average of the three maximum voluntary contractions were used for the analysis.²⁵ The pressure values were categorized as very weak for pressure values ranging from 7.5 to 14.5 cmH₂O; weak for pressure values from 14.6 to 26.5 cmH₂O; moderate for pressure values varying from 26.6 to 41.5 cmH₂O; good for pressure values from 41.6 to 60.5 cmH₂O; and strong for pressure values at or above 60.6 cmH₂O.²⁶

Data analyses

Data were analyzed by the Statistical Package for Social Sciences Software (SPSS) 20.0 for Windows. Continuous data are presented as mean \pm standard deviation, after being tested for normality with the Kolmogorov-Smirnov test. Categorical variables are presented as counts and percentages. Paired Student's t-tests were used to compare the thermographic analyses between rest and during the contractions. Pearson's correlation was used to analyze the thermographic and vaginal manometry data. A value of $p < 0.05$ was set as statistical significance.

Results

Two-hundred and fifty-nine women was screened for eligibility. Nineteen individuals did not perform an effective PFM contraction, and nine felt discomfort during the probe introduction. Two-hundred and thirty-one women completed the study. Table 1 provides the sociodemographic and gynecology-obstetric information of the participants.

Considering the thermographic characteristics at rest and during maximum voluntary contraction, the color images showed a greater percentage of white and red colors (classified as warm) during contraction (Fig. 1). Minimum, average, and maximal temperatures of the PFM were not different when comparing at rest and during contraction (Table 2).

Table 1 Sociodemographic, clinical, and gynecology-obstetric descriptive data.

Variables	n = 231
Age ^a (years)	58.4 \pm 6
Body temperature ^a °C	36 \pm 0.3
Marital Status (%)	
With partner	71
Without partner	29
Parity ^a	2 \pm 1
Types of delivery (%)	
Vaginal	39
Cesarean section	32
Vaginal e cesarean section	29
Menstrual Cycle (%)	
Premenopause	31
Peri menopause	32
Postmenopause	28
Hysterectomy	9
Body mass index (%)	
Normal	27
Overweight	44
Obesity	29
Manometry (%)	
Very weak	4
Weak	23
Moderate	67
Good	5
Strong	1

^a Results are mean \pm standard deviation and percentage.

The percentages of white, red, orange, yellow, green, cyan, and blue colors were different between rest and during contraction (Table 2). There was a positive and strong correlation between PFM pressure and PF average temperature (Table 3).

Discussion

This study shows novel data about assessment of the PFM by using IRT. This method was able to identify color differences between the PFM at rest and during PFM maximal voluntary contraction. A positive and strong correlation between PF temperature and PFM squeeze pressure was also found, and indicates that the higher the temperature the greater the PFM squeeze pressure. In fact, during muscle contraction, energy generated is dissipated and conducted to the skin through blood flow.^{27,28}

In this study, warm and cold colors during the evaluation of the PF by thermography had different patterns. For example, colors considered colder filled approximately 56% of the image at rest. Conversely, during muscle contraction, warmer colors filled nearly 51% of the image. This may occur because phasic fibers in the PF are located more centrally and superficially. These fibers are the first to be triggered during muscle contraction and their main feature is to quickly generate a large amount of force,^{29,30} which means that muscle anatomic location and its physiology are variables that can affect thermographic results. It is important to

Table 2 Thermographic characteristics of the pelvic floor region at rest and during pelvic floor muscle contraction.

	PFM at rest	PFM contraction	Mean difference (95% Confidence Interval)
Minimum temperature (°C)	33.8 ± 0.5	33.8 ± 1.0	0.0 (-0.1, 0.1)
Average temperature (°C)	36.4 ± 0.8	36.4 ± 0.8	0.0 (-0.1, 0.0)
Maximum temperature (°C)	37.6 ± 0.5	37.6 ± 0.9	0.0 (0.0, 0.1)
White	1.0 ± 0.1	2.1 ± 0.2	-1.1 (-1.2, -1.1)
Red	15.8 ± 0.8	17.4 ± 1.0	-1.5 (-1.7, -1.4)
Orange	16.4 ± 0.7	18.0 ± 0.8	-1.6 (-1.8, -1.5)
Yellow	10.1 ± 0.8	13.9 ± 1.5	-3.7 (-3.9, -3.5)
Green	24.9 ± 0.8	20.8 ± 1.3	4.0 (3.8, 4.2)
Cyan	6.1 ± 0.3	6.7 ± 0.3	-0.6 (-0.6, -0.5)
Blue	25.2 ± 1.0	20.5 ± 1.6	4.7 (4.5, 5.0)
Violet	0	0	0.0 (0.0, 0.0)
Magenta	0.2 ± 0.1	0.2 ± 0.1	0.0 (0.0, 0.0)

Results in percentage (mean ± standard deviation); °C: degree Celsius.

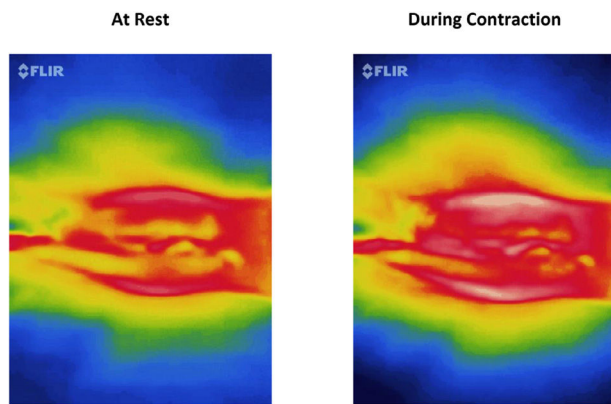


Fig. 1 Infrared thermography of a single woman at rest and during muscle contraction. It shows an increasing amount of red color in the center of the image during pelvic floor muscles contraction.

mention that there are no studies that performed a qualitative investigation between temperature and muscle function. Research on muscle temperature mainly concerns the association of issues and environment or muscle temperature on fatigue and strength.³¹

Although there was a difference among the colors' distributions, no differences were found in the temperature values (minimum, average, and maximum) between PFM at rest and during contraction. This finding is probably related

to the fact that PFM contractions performed in this study were not a physical exercise protocol. Therefore, it is possible that in the absence of strength training the force generated was insufficient to generate a local temperature change.²⁹ Some studies in this area used muscle load to evaluate fatigue.³² It means that the more the force exerted during tests, the greater the changes in the temperature pattern. So, the physiologic understanding of these studies cannot be associated with researches that assessed only three voluntary muscle contractions.^{33,34}

Our results showed a strong correlation between PFM pressure and the PF temperature during maximum voluntary contraction. Formenti et al.³⁵ reported an increase in skin temperature during anaerobic exercise and a decrease after the exercise, associating these differences to muscular power. It is important to emphasize that skin temperature measured by the IRT depends on several factors, such as the environment, heat dissipation,^{36,37} and the previous level of strength.³⁸ In this study, as the environment and heat dissipation were controlled, the levels of strength might have been the main aspect associated with this strong correlation.

We aimed to understand the IRT pattern of the PFM at rest and during muscular contraction. The literature shows that the use of thermography helps to understand the physiological mechanisms that occur during exercise.³⁹ Vasoconstriction of the skin is expected early in exercise due to an increase of blood flow to active muscles during contraction. However, during resistance exercise, central temperature regulatory mechanisms promote cutaneous vasodilation, so

Table 3 Correlation between PFM pressure and average temperature of the PF area.

	n	Manometry (cmH ₂ O)	PF temperature (°C)	Correlation	p value
Very weak	9	9.5 ± 2.7	34.9 ± 0.7	0.7	0.001*
Weak	53	19.3 ± 2.8	35.6 ± 0.8		
Moderate	154	34.5 ± 3.2	36.7 ± 0.5		
Good	12	53.7 ± 5.6	37.5 ± 0.1		
Strong	3	63.2 ± 1.4	37.6 ± 0.0		

Results in mean ± standard deviation; n, number of participants; manometry, PFM pressure recorded by the manometer; °C, PF average temperature in degree Celsius; PFM, pelvic floor muscles; PF, pelvic floor.

* Significant by Pearson's correlation.

heat dissipation occurs through the skin. This mechanism is called thermoregulation, which removes excessive body heat, allowing the individual to maintain exercise longer.^{39,40} For these reasons, we believe that thermography can be used as another resource for PFM evaluation, with potential of adjusting the intensity of the training protocols, in addition to evaluating their effectiveness.

We consider thermography to be a potential tool for PFM analysis, due to the possibility of its use in cases in which intracavitary evaluation is limited or not possible, for example, postpartum in the presence of physiological lochia or pain. Moreover, when detecting temperature alterations by using IRT, the health sciences have considered this imaging exam to be a diagnostic tool to reduce the exposure of individuals to ionizing radiation.⁴¹ However, comparisons of IRT to other diagnostic exams that use radiation to assess the PF have not been documented yet. The FLIR ONE is recommended because it is feasible, mobile, and provides fast measurements in clinical burn practice; nonetheless, its routine use in evaluating the PF requires additional research.³⁸

Finally, it is important to mention that a sample size calculation was not performed in this study and only 45- to 70-year-old women were included, thus limiting the external validity of the results. However, IRT as presented in this study is relevant due to its wide clinical applicability.

Conclusion

IRT was able to identify differences in colors at rest and during PFM maximum voluntary contraction. However, there was no difference in overall PF temperature. White, red, orange, or yellow colors became more visible in the center of the image during contraction. Also, there was a strong correlation between vaginal manometry and the temperature of the PF area.

Conflicts of interest

None to declare.

Acknowledgements

This study was financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brazil (CAPES) - Finance Code 001.

References

- Milsom I, Gyhagen M. The prevalence of urinary incontinence. *Climacteric*. 2019;22:217–222.
- Ng KS, Sivakumaran Y, Nassar N, Gladman MA. Fecal Incontinence: community prevalence and associated factors. A systematic review. *Dis Colon Rectum*. 2015;58:1194–1209.
- McCabe MP, Sharlip ID, Lewis R, et al. Incidence and prevalence of sexual dysfunction in women and men: a consensus statement from the fourth international consultation on sexual medicine 2015. *J Sex Med*. 2016;13:144–152.
- Chevalier F, Fernandez-Lao C, Cuesta-Vargas AI. Normal reference values of strength in pelvic floor muscle of women: a descriptive and inferential study. *BMC Womens Health*. 2014;14:143.
- Bø K, Sherburn M. Evaluation of female pelvic-floor muscle function and strength. *Phys Ther*. 2005;85(3):269–282.
- Scheidt S, Rüwald J, Schildberg FA, et al. A systematic review on the value of infrared thermography in the early detection of periprosthetic joint infections. *Z Orthop Unfall*. 2019.
- Singh D, Singh AK. Role of image thermography in early breast cancer detection- past, present and future. *Comput Methods Programs Biomed*. 2020;183: 105074.
- Vardasca R, Magalhaes C, Silva P, Abreu P, Mendes J, Restivo MT. Biomedical musculoskeletal applications of infrared thermal imaging on arm and forearm: a systematic review. *J Therm Biol*. 2019;82:164–177.
- Côrte AC, Pedrinelli A, Marttos A, Souza I, Grava J, Hernandez A. Infrared thermography study as a complementary method of screening and prevention of muscle injuries: pilot study. *BMJ Open Sport Exerc Med*. 2019;5(1) e000431.
- Ng EYK. A review of thermography as promising non-invasive detection modality for breast tumor. *Int J Thermal Sci*. 2009;48(5):849–859.
- Hildebrandt C, Raschner C, Amme K. An overview of recent application of medical infrared thermography in sports medicine in Austria. *Sensors (Basel)*. 2010;10:4700–4715.
- Love TJ. *Analysis and Application of Thermography in Medical Diagnosis. Heat Transfer in Medicine and Biology*. Boston, MA: Springer; 1985:333–352.
- van Lunsen RH, Laan E. Genital vascular responsiveness and sexual feelings in midlife women: psychophysiological, brain, and genital imaging studies. *Menopause*. 2004;11:741–748.
- Sancibrian R, Gutierrez-Diez MC, Redondo-Figuero C, Llata JR, Manuel-Palazuelos JC. Using infrared imaging for assessment of muscular activity in the forearm of surgeons in the performance of laparoscopic tasks. *Proc Inst Mech Eng H*. 2019;233: 999–1009.
- Gómez-Carmona P, Fernández-Cuevas I, Sillero-Quintana M, Arnaiz-Lastras J, Navandar A. Infrared thermography protocol on reducing the incidence of soccer injuries. *J Sport Rehabil*. 2020;29(8):1222–1227.
- von Elm E, Altman DG, Egger M, et al. The strengthening the reporting of observational studies in epidemiology (STROBE) statement: guidelines for reporting observational studies. *J Clin Epidemiol*. 2008;61(4):344–349.
- Persu C, Chapple CR, Cauni V, Gutue S, Geavlete P. Pelvic organ prolapse quantification system (POP-Q) - a new era in pelvic prolapse staging. *J Med Life*. 2011;4(1):75–81.
- Fernández-Cuevas I, Marins JCB, Lastras JA, et al. Classification of factors influencing the use of infrared thermography in humans: a review. *Infrared Phys Techn*. 2015;71:28–55.
- Fernández-Cuevas I, Marins JCB, Lastras JA, et al. Classification of factors influencing the use of infrared thermography in humans: a review. *Infrared Phys Techn*. 2015;71:28–55.
- Umapathy S, Thulasi R, Gupta N, Sivanadhan S. Thermography and colour Doppler ultrasound: a potential complementary diagnostic tool in evaluation of rheumatoid arthritis in the knee region. *Biomed Tech (Berl)*. 2020;65:289–299.
- Nazzaro G, Moltrasio C, Marzano AV. Infrared thermography and color Doppler: two combined tools for assessing inflammation in hidradenitis suppurativa. *Skin Res Technol*. 2020;26:140–141.
- Lerkvaleekul B, Jaovisidha S, Sungkarat W, et al. The comparisons between thermography and ultrasonography with physical examination for wrist joint assessment in juvenile idiopathic arthritis. *Physiol Meas*. 2017;38:691–700.
- Baek SW, Lee JM, Park YB, Park YJ. Relationship between tongue temperature estimated by infrared thermography, tongue color, and cold-heat pathological patterns: a

- retrospective chart review study. *Evid Based Complement Alternat Med*. 2018;6841460.
24. Ferreira CH, Barbosa PB, de Oliveira Souza F, Antônio FI, Franco MM, Bø K. Inter-rater reliability study of the modified Oxford Grading Scale and the Peritron manometer. *Physiotherapy*. 2011;97:132–138.
 25. Pereira VS, Hirakawa HS, Oliveira AB, Driusso P. Relationship among vaginal palpation, vaginal squeeze pressure, electromyographic and ultrasonographic variables of female pelvic floor muscles. *Braz J Phys Ther*. 2014;18:428–434.
 26. Angelo PH, Varella LRD, de Oliveira MCE, et al. A manometry classification to assess pelvic floor muscle function in women. *PLoS One*. 2017;12: e0187045.
 27. Cramer MN, Jay O. Biophysical aspects of human thermoregulation during heat stress. *Auton Neurosci*. 2016;196:3–13.
 28. Vargas JVC, Brioschi ML, Dias FG, et al. Normalized methodology for medical infrared imaging. *Infrared Phys Technol*. 2009;52(1):42–47.
 29. Eickmeyer SM. Anatomy and physiology of the pelvic floor. *Phys Med Rehabil Clin N Am*. 2017;28:455–460.
 30. Rocca Rossetti S. Functional anatomy of pelvic floor. *Arch Ital Urol Androl*. 2016;88:28–37.
 31. Bartuzi P, Roman-Liu D, Wiśniewski T. The influence of fatigue on muscle temperature. *Int J Occup Saf Ergon*. 2012;18:233–243.
 32. Teng M, Kervinio F, Moutounaïck M, et al. Review of pelvic and perineal neuromuscular fatigue: evaluation and impact on therapeutic strategies. *Ann Phys Rehabil Med*. 2018;61:345–351.
 33. Neves EB, Vilaca-Alves J, Antunes N, Felisberto IM, Rosa C, Reis VM. Different responses of the skin temperature to physical exercise: systematic review. *Conf Proc IEEE Eng Med Biol Soc*. 2015;2015:1307–1310.
 34. Elvin A, Siösteen AK, Nilsson A, Kosek E. Decreased muscle blood flow in fibromyalgia patients during standardised muscle exercise: a contrast media enhanced colour Doppler study. *Eur J Pain*. 2006;10:137–144.
 35. Formenti D, Ludwig N, Gargano M, et al. Thermal imaging of exercise-associated skin temperature changes in trained and untrained female subjects. *Ann Biomed Eng*. 2013;41: 863–871.
 36. Gisolfi CV, Wenger CB. Temperature regulation during exercise: old concepts, new ideas. *Exerc. Sport Sci. Rev.*. 1984;12:399–416.
 37. Schlager O, Gschwandtner ME, Herberg K, et al. Correlation of infrared thermography and skin perfusion in Raynaud patients and in healthy controls. *Microvasc Res*. 2010;80:54–57.
 38. Hadžić V, Širok B, Malnersić A, Čoh M. Can infrared thermography be used to monitor fatigue during exercise? A case study. *J Sport Health Sci*. 2019;8:89–92.
 39. Hadžić V, Širok B, Malnersić A, Čoh M. Can infrared thermography be used to monitor fatigue during exercise? A case study. *J Sport Health Sci*. 2019;8:89–92.
 40. Weigert M, Nitzsche N, Kunert F, Lösch C, Schulz H. The influence of body composition on exercise-associated skin temperature changes after resistance training. *J Therm Biol*. 2018;75: 112–119.
 41. Sanchis-Sánchez E, Vergara-Hernández C, Cibrián RM, Salvador R, Sanchis E, Codoñer-Franch P. Infrared thermal imaging in the diagnosis of musculoskeletal injuries: a systematic review and meta-analysis. *AJR Am J Roentgenol*. 2014;203:875–882.